Protocol for Hepatic (Liver) Tissue Homogenization in the Bullet Blender™

The protocol described in this document is for the use of the Bullet Blender $^{\text{TM}}$ for the homogenization of liver / hepatic tissue (from a variety of animals). Note that the time and speed settings may differ due to the variation in consistency / texture of liver tissue from species to species. This protocol does not specify a particular buffer - you may choose which is most appropriate for your downstream application (nucleic acid isolation, protein extraction, etc.).

Materials Required:

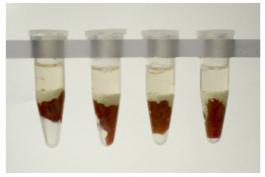
liver tissue, saline, Bullet Blender™, homogenization buffer, pipettor, microcentrifuge tubes, and <u>1.0mm zirconium oxide</u> beads or 0.1mm glass beads

Instructions

- **1.** Cut liver into appropriately sized pieces for analysis (50mg-300mg) and place into a microcentrifuge tube.
- 2. **OPTIONAL:** Wash tissue 3x with ~1mL PBS. Aspirate. **NOTE:** This step removes external contaminants (blood, etc.).
- **3.** Add zirconium oxide beads (1.0mm) **OR** glass beads (0.1mm) to the tube. Use a mass of beads equal to your mass of tissue.
- **4.** Add 0.1mL to 0.6mL buffer (2 volumes of buffer for every volume of cells).
- **5.** Close the microcentrifuge tubes.
- **6.** Place tubes into the Bullet Blender[™].
- 7. Set controls for **SPEED 8** and **TIME 3** minutes. Press **Start**.
- **8.** After the run, remove tubes from the instrument.
- **9.** Visually inspect samples. If homogenization is unsatisfactory, run for another two minutes at the **SPEED 8.**
- **10.** Proceed with your downstream application.

SAFETY NOTE!!!

When using a centrifuge to separate your homogenate from the debris and beads, make sure your tubes are balanced.





before

after



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